



## **UWL REPOSITORY**

**repository.uwl.ac.uk**

Comparative genomics analysis demonstrated a link between staphylococci isolated from different sources: a possible public health risk

Cave, Rory J., Misra, Raju, Chen, Jiazhen, Wang, Shiyong and Mkrtyan, Hermine (2021) Comparative genomics analysis demonstrated a link between staphylococci isolated from different sources: a possible public health risk. *Frontiers in Microbiology*, 12.

<http://dx.doi.org/10.3389/fmicb.2021.576696>

This is the Accepted Version of the final output.

UWL repository link: <https://repository.uwl.ac.uk/id/eprint/7636/>

**Alternative formats:** If you require this document in an alternative format, please contact: [open.research@uwl.ac.uk](mailto:open.research@uwl.ac.uk)

**Copyright:** Creative Commons: Attribution 4.0

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

**Take down policy:** If you believe that this document breaches copyright, please contact us at [open.research@uwl.ac.uk](mailto:open.research@uwl.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.

# Comparative genomics analysis demonstrated a link between staphylococci isolated from different sources: A Possible public health risk.

Rory Cave<sup>1</sup> Raju Misra<sup>2</sup>; Jiazhen Chen<sup>3</sup>; Shiyong Wang<sup>3</sup>; Hermine V Mkrtchyan<sup>4,\*</sup>

<sup>1</sup>University of East London, School of Health, Sport and Bioscience, Water Lane, London,

<sup>2</sup>Natural History Museum, Core Research Laboratories, Molecular Biology, Cromwell Rd, London

<sup>3</sup>Department of Infectious Disease, Huashan Hospital, Fudan University, 12 Middle Wulumuqi Rd., Shanghai 200040, China

<sup>4</sup>School of Biomedical Sciences, University of West London, St Mary's Rd, London, UK, W5 5RF

## \* Correspondence:

Hermine V Mkrtchyan

[hermine.mkrchyan@uwl.ac.uk](mailto:hermine.mkrchyan@uwl.ac.uk)

## Public settings, WGS, Phylogenetic, Staphylococci, public health

### Abstract

Coagulase-negative staphylococci (CoNS) have been recovered from different ecological niches, however, little is known about the genetic relatedness of these isolates. In this study, we used whole genome sequencing to compare *mecA* positive (*mecA*<sup>+</sup>) *Staphylococcus epidermidis*, *Staphylococcus haemolyticus* and *Staphylococcus hominis* isolates recovered from hand-touched surfaces from general public settings in East and West London with data of isolates deposited to European Nucleotide Archive (ENA) by other research groups. These included isolates associated with hospital settings (including those recovered from patients), healthy humans, livestock, pets, plants and natural and other public environments. Using core and accessory phylogenetic analyses we were able to identify that the *mecA*<sup>+</sup> *S. epidermidis* and *S. haemolyticus* isolates recovered from general public settings were genetically related to isolates recovered from the bloodstream, urinary tract and eye infections. *S. epidermidis* isolates recovered in our study were also shown to be genetically related to isolates previously recovered from livestock/livestock housing, whereas *S. haemolyticus* isolates were genetically related to isolates recovered from a dog and kefir (fermented cow milk drink). *MecA*<sup>+</sup> *S. hominis* isolates were not genetically related to any isolates recovered from clinical samples but were genetically related to isolates recovered from mosquitoes, air samples (residential areas) and kefir. All three species showed to have genetic relatedness to isolates recovered from healthy humans. These results show that CoNS isolates in this study share genetic similarities with those of different lineages and that *mecA*<sup>+</sup> *S. epidermidis* and *S. haemolyticus* isolates found in general public settings in this study may pose a risk to public health.

## 39 Introduction

40 Hand touched surfaces in public areas act as an intermediate for human to human transmission  
41 of pathogenic bacteria (Lei et al., 2017). Many bacteria **responsible for various infections are**  
42 known to have originated from humans or animals' sources which later transmitted across the  
43 species barrier (Fitzgerald, 2012; Argudín et al., 2015b) However, little is known of the genetic  
44 lineages of coagulase-negative staphylococci (CoNS) recovered from hand touched surfaces in  
45 public settings and the threat they pose to public health.

46 CoNS are the most common commensal group of bacteria found on human skin and frequently  
47 found on surfaces in hospitals, hotel rooms, libraries, university campus and public transport  
48 (Becker et al., 2014; Seng et al., 2017; Xu et al., 2018b; Cave et al., 2019). Unlike coagulase-  
49 positive *Staphylococcus aureus* they lack many key virulent factors; however, *Staphylococcus*  
50 *epidermidis*, *Staphylococcus haemolyticus* and *Staphylococcus hominis* have been identified  
51 as significant pathogens associated with nosocomial infections and medical devices (Chaves et  
52 al., 2005; Cherifi et al., 2013; Czekaj et al., 2015). *S. epidermidis* accounts for 22% of  
53 bloodstream infections in intensive care unit patients in the USA, whereas CoNS accounts for  
54 23.1% and 12.7% of bloodstream infections in Israel and China respectively (Otto, 2009; Abu-  
55 Saleh et al., 2018; Cui et al., 2019). In addition, CoNS have also been reported to be associated  
56 with community and animal/livestock infections (Nanoukon et al., 2017; Schoenfelder et al.,  
57 2017). Many CoNS infections are challenging to treat due to them being resistant to multiple  
58 antibiotics (Lee et al., 2018; Cui et al., 2019; Pain et al., 2019). One **antibiotic resistance gene**  
59 in particular which **makes infections caused by staphylococci** challenging to treat is *mecA*. This  
60 gene confers resistance to **all beta-lactam antibiotics**, including carbapenem, cephalosporin,  
61 penam, cephamycin and monobactam and is associated with the “superbug” methicillin  
62 resistant *Staphylococcus aureus* (MRSA), a **bacterium** that has caused severe infections in  
63 **healthcare settings**, community and livestock worldwide.

64 London is the most **densely** populated city in Europe, with a population of 8.7 million people  
65 (2016) and a population density of 5,590 per km<sup>2</sup> (2016) (Land Area and Population Density,  
66 Ward and Borough - London Datastore). In 2018, it was estimated that there were 60,788 cases  
67 of antibiotic-resistant severe infections in England with London having the highest rate of  
68 bloodstream infections **caused by antibiotic resistant bacteria** (42.9 per 100,000 population)  
69 (English Surveillance Programme for Antimicrobial Utilisation and Resistance (ESPAUR),  
70 2019). In addition, CoNS can be commonly isolated from environmental sites in London; with  
71 one study showing that 94% of the recovered CoNS isolates were antibiotic-resistant and 11%  
72 carrying the *mecA* gene (Xu et al., 2018b). Currently, due to the improvement of whole genome  
73 sequencing (WGS) technology, it is possible to determine the genetic relationship of less  
74 frequently studied bacteria from different settings (Quainoo et al., 2017).

75 In this study, we genetically **compared** *mecA* positive (*mecA*<sup>+</sup>) *S. epidermidis*, *S. haemolyticus*  
76 and *S. hominis* **isolates** recovered from high-frequency touched surfaces of general public  
77 settings in the community and public areas in hospitals with complete and draft genomes  
78 (obtained from the European Nucleotide Archive (ENA)) of the isolates recovered from  
79 different sources, including bloodstream, urinary tract, and eye infections, healthy humans,  
80 livestock, pets, plants, fermented **milk drink**, natural and other public environmental **sites**. **This**  
81 **comparative genomics analysis helped to assess whether *mecA*<sup>+</sup> CoNS isolates recovered in**  
82 **our study were genetically similar to isolates recovered from different sources (obtained from**  
83 **the ENA), including those that have previously been reported to cause infections.**

## 84 Materials and Methods

## 85 **Sample collection**

86 Staphylococcal isolates were recovered between November 2016 to September 2017 from  
87 high-frequency hand touched surfaces of inanimate objects from two locations in East London  
88 and two locations in West London. These locations included one area considered as public  
89 settings (shopping centres and train stations) and another area within hospitals where the  
90 general public had easy access, without being a patient (reception area, public washrooms,  
91 corridors, lifts). Fifty sites from each location were randomly sampled using COPAN dry  
92 swabs (Copan Diagnostics Inc., USA). Six hundred staphylococcal isolates were recovered of  
93 which 224 were from East London and 376 from West London. One hundred and eighty-two  
94 of the isolates were from the community area and 418 from hospital areas. Ninety-seven of the  
95 isolates were from East London community area; 85 from West London Community area; 127  
96 from **East London Hospital (ELH)** and 376 from **West London Hospital (WLH)** (Cave et al.,  
97 2019).

98

## 99 **Isolation of staphylococci**

100 All samples were directly inoculated onto mannitol salt agar (MSA, Oxoid Basingstoke, UK)  
101 within 1-3 hours of recovery and incubated aerobically for 24-72 hours at 37°C. To prevent  
102 bias up to 10 colonies from each plate were picked each having different colony morphology  
103 or if there are less than 10 different colony morphologies an equal amount of different colony  
104 morphologies was selected. These isolates were screened for potential staphylococci  
105 characteristics, including performing catalase and coagulase tests. Prolex™ staph latex kit  
106 (ProLab Diagnostics, Neston, UK) was used to distinguish *S. aureus* and coagulase-negative  
107 staphylococci (Cave et al., 2019).

108

## 109 **Identification of staphylococci recovered from high-frequency hand touch areas**

110 Staphylococcal isolates were initially identified by gram staining and catalase test. The  
111 presumptive staphylococcal isolates were further identified at species level using Matrix-  
112 assisted laser desorption ionisation-time of flight mass spectroscopy (MALDI-TOF-MS,  
113 Microflex LT, Bruker Daltonics, Coventry, UK) in a positive linear mode (2000–20,000 m/z  
114 range). Samples were prepared using a full protein extraction method as described previously.  
115 (Cave et al., 2019).

116

## 117 **Detection of the *mecA* gene by PCR**

118 Forty three bacterial isolates DNA was extracted via boil lysis **and used to perform PCR for**  
119 **detection of** the *mecA* gene as previously described (Cave et al., 2019). Using the Met1 and  
120 Met2 primers (Eurofins, Germany) PCR reactions were performed in a 20 µl volume (Hanssen  
121 et al., 2004), **consisting** of 10 µl of Phusion Master Mix; 1 µl of forward primer, 1 µl of reverse  
122 primer, 7 µl of sterile distilled water and 1 µl of isolates DNA template. The PCR condition  
123 for this reaction was 94 °C for 5 minutes followed by 35 cycles of denaturation at 94 °C for 30  
124 seconds, annealing at 52 °C for 30 seconds and extension at 72 °C for 1 minute with a final  
125 extension at 72 °C for 10 minutes.

126

## 127 **Genome sequencing and assembly**

128 Whole genome sequencing was performed for 43 *mecA*<sup>+</sup> staphylococci isolates using Illumina  
129 HiSeq platform. Seven out of 43 isolates were whole genome sequenced by MicrobesNG  
130 (Birmingham, UK) and the remaining isolates were sequenced at Fudan University, Shanghai,  
131 China (Supplementary Table S1).

132 Genomic DNA was extracted using TIANamp Bacteria DNA kit (Tiangen, China) and paired-  
133 end sequencing libraries were constructed using Nextera XT DNA Sample Preparation kits or  
134 TruSeq DNA HT Sample Prep Kit (Illumina, USA) following manufacturer's instruction.

135 Quality of reads was assessed using FASTQC and trimmed using trimmomatic (Version 0.35),  
136 default settings, specifying a Phred cutoff of Q20 (Andrews, 2011; Bolger et al., 2014).  
137 Trimmed reads were *de novo* assembly by SPAdes 3.11 and contig assembly was analysed by  
138 QUAST and contigs that were  $\leq 500$ bp were removed (Bankevich et al., 2012; Gurevich et al.,  
139 2013). The species of these isolates were confirmed by 16s RNA sequencing from the  
140 assembled genomes by using the barrnap software (<https://github.com/tseemann/barrnap>) and  
141 searched against a database of known 16S ribosomal RNA sequences using NCBI BLAST tool  
142 with a cutoff for species identity of 95% similarity (Altschul et al., 1990; Janda and Abbott,  
143 2007).

#### 144 **Phylogenetic analyses**

145 A core SNP Maximal likelihood tree was constructed using isolates previously recovered from  
146 different sources (data obtained from the ENA, [accessed in July 2019; Supplementary Table](#)  
147 [S2](#)). [The main selection criteria of the isolates obtained from ENA database was their isolation](#)  
148 [sources, which have been recorded in the ENA and/or reported in a peer-reviewed literature.](#)  
149 [The selection criteria for isolates recovered in this study was the presence of clinically](#)  
150 [important \*mecA\* gene.](#) SMALT version 0.5.8 (<https://www.sanger.ac.uk/science/tools>) was  
151 used to map short reads against reference genomes. Reference genomes used to map each  
152 staphylococcal species included *S. epidermidis* ATTC 11228; *S. haemolyticus* JCSC 1435 and  
153 *S. hominis* K1. SNP calling was done in parallel with all samples of the same species using  
154 VarScan version 2.3.9 (Koboldt et al., 2009). VCF file was converted to multi-FASTA  
155 alignment file using the python script [vcf2phylip](#)  
156 (<https://github.com/edgardomortiz/vcf2phylip>). Recombination was detected and removed  
157 from the genome with Gubbins (Croucher et al., 2015). A maximal likelihood tree was  
158 constructed using RAxML version 8 using the generalised time reversible model (GTR) model  
159 with GAMMA method of correction for site rate variation and 100 bootstrap replications  
160 (Stamatakis, 2014). The phylogenetic tree was visualised and annotated using ITOL (Letunic  
161 and Bork, 2016).

162 The distance of the accessory genome for each isolate was determined using the variable-length  
163 *k*-mer comparisons to distinguish isolates' divergence in shared sequence and gene content  
164 using the POPpunk pipeline (Lees et al., 2019). [The number of mixture components was](#)  
165 [adjusted for each species for obtaining a low-density score \(proportion of edges in the](#)  
166 [network\), high transitivity score and high overall score \(network score based on density and](#)  
167 [transitivity\).](#) Accessory genome distance was determined by t-SNE with the perplexity (the  
168 [number of close neighbours each point has](#)) was adjusted for each species to provide the  
169 [clearest picture of clustering and visualised using Microreact \(Argimón et al., 2016\).](#)

170 **Identifying the unique accessory genes that were only present in the ENA *S. hominis***  
171 **clinical isolates**

172 To determine the unique accessory genes in *S. hominis* hospital associated isolates obtained  
173 from the ENA, pangenome analysis was performed using Roary pipeline version 3.4.2. We  
174 then compared the differences in genes between all *S. hominis* hospital associated isolates  
175 obtained from the ENA with the rest of the *S. hominis* isolates used in phylogenetic analyses  
176 (Page et al., 2015).

## 177 **Results**

### 178 **CoNS *mecA* from public settings**

179 The *mecA* gene was identified in 43 isolates that were recovered from high frequency touched  
180 surfaces from general public settings from the community and public areas in hospitals (Table  
181 1). This included: *S. epidermidis* (n=17); *S. haemolyticus* (n=10), *S. hominis* (n=10), *S. cohnii*  
182 (n=3) and *S. warneri* (n=3).

### 183 **Phylogenetic analysis of *S. epidermidis***

184 Phylogenetic analysis was performed to determine the relatedness of environmental isolates in  
185 this study with those recovered from other sources, including isolates recovered from infections  
186 (ENA). In this study, 17 *S. epidermidis mecA*<sup>+</sup> isolates recovered from East and West London  
187 were compared to those obtained from the ENA that have previously been recovered from  
188 infections (n=34); healthy humans (n=9), livestock (n=13) rodents (n=2), plants (n=4), hospital  
189 environment (n=7), animal housing environments (n=2) and natural environments (n=2). Core  
190 single nucleotide polymorphism (SNP) phylogenetic tree analysis identified two distinctive  
191 clades of which 59 out of 90 were *mecA*<sup>+</sup> (Figure 1). Four isolates from East London Hospital  
192 (ELH) (321, 327, 329 and 355) belonged to clade A, whereas 1 isolate (407) recovered from  
193 ELH was in clade B together with all (n= 12) isolates recovered from **West London Hospital**  
194 **(WLH)**. Interestingly, all ENA isolates recovered from infections, except for VCU128 which  
195 was recovered from human airways, were found to be in clade B. ENA isolates recovered from  
196 healthy humans, and animals were found within both clades whereas those recovered from  
197 plants (all rice seeds) were found in clade A only. Isolate 355 recovered from public settings  
198 in our study was genetically related to those ENA isolates that have been recovered from  
199 healthy humans (MRSE 52-2 and NIHLM057); isolate 407 was genetically related to those  
200 isolates that have been recovered from cow (Y24), pig (PR246B0) and animal housing (M01  
201 and M025), whereas isolates **435, 436, 465, 475, 631, 673, 711, 712, 713, 715 and 716 from**  
202 **WLH** were genetically related to isolates previously recovered (ENA) from bloodstream  
203 (B45679, 764 SEPI, FDARGOS 153, FDARGOS 83, VCU045, SH06 17, SH03 17, SH03 17)  
204 and an endotracheal tube biofilm of a mechanically ventilated patient (ET-0240). 321, 327 and  
205 329 recovered from ELH uniquely showed no relatedness to any other isolate.

206 PopPUNK analyses revealed that *S. epidermidis* isolates can be combined into 31 groups by their  
207 combined core and accessory genome. The accessory genome t-SNE analyses, set at the perplexity of  
208 20, showed that there were 5 distinct clusters (Figure 2). Two of these groups had a mixture of isolates  
209 belonging to different combined clusters. None of the clusters includes isolates belonging to a single  
210 sequence type. In this study, the *mecA*<sup>+</sup> *S. epidermidis* isolates recovered from East and West London  
211 were found in different clusters. The accessory genome of *mecA*<sup>+</sup> *S. epidermidis* isolates recovered  
212 from public settings was related to those ENA isolates that have been previously recovered from  
213 bloodstream infections (B45679-10, FDAARGOS 83 and FDAARGOS 161), infected airways  
214 (VCU45 and VCU128) and cerebrospinal fluid (CSF41498), endotracheal tube biofilm of a  
215 mechanically ventilated patient (ET-024), central venous catheter (1457); from healthy human skin  
216 (14.1.R1, mucosa (ATCC 12228) and human airways (MRSE 52-2); from livestock (cows (SNUC 901,  
217 SNUC 3608, SNUC 75, SNUC, y24, PM221 and NW32), (a pig (PR246B0 and a sheep (AG42)), a  
218 mouse (SCL25); plants (SE2.9, SE4.8, SE4.7 and SE4.6), and from a natural environment (SNUT). In  
219 addition, we found that cluster 1 included ENA isolates that were recovered from hospital  
220 environments in medical wards and isolates recovered from bloodstream infections, whereas cluster 4  
221 included isolates all recovered from the bloodstream and only a single isolate from healthy human skin  
222 (M008).

### 223 **Phylogenetic analyses of *S. haemolyticus***

224 Ten *S. haemolyticus mecA*<sup>+</sup> isolates recovered from public settings in East and West London were  
225 phylogenetically compared with isolates recovered from other sources, including those recovered from  
226 different infections (n=48) and central venous catheter (n=2), commensal isolates recovered from  
227 human skin, nares and eyes (n=7); livestock (cow; n=7), kefir (fermented cow milk drink; n=1),  
228 companion animals (dog; n=1), public settings (from the surface area of a building and tropical air  
229 sample (n=2)), natural environments (n=3), and plant-associated isolates (n=2). The core SNP  
230 maximum likelihood phylogenetic tree revealed two distinctive clades (Clade A and Clade B), of which  
231 65 out of 83 isolates were identified as *mecA*<sup>+</sup> (Figure 3). Clade A consisted of ENA isolates included  
232 those recovered from the bloodstream, vagina and sputum, livestock (cows), a companion animal  
233 (dog), groundwater and healthy human eyes. Clade B consisted of ENA isolates recovered from the  
234 bloodstream, eye infection, colon infection, central venous catheter, healthy human skin, kefir, willow  
235 tree, livestock (cows), tropical air samples, copper alloy coin, surface area of a building and waste and  
236 the hygiene compartment of the International Space Station. In addition, all *mecA*<sup>+</sup> *S. haemolyticus*  
237 isolates recovered from public areas in London in our study were found in clade B, except for the  
238 isolate 492, which was recovered from (WLH). We found that isolates we recovered from public areas  
239 in ELH and WLH (373, 445 and 538) and isolates recovered from the public settings in East London  
240 (1, 93, 99 and 105) were genetically related to ENA isolates that have previously been recovered from  
241 an eye infection (SH1572), bloodstream (M-176), and central venous catheter (95671). In addition, one  
242 isolate recovered from public areas in WLH (492) in this study were genetically related to an ENA  
243 isolate that has previously been recovered from a dog (SW007), whereas one isolate (445) was  
244 genetically similar to an isolate that was recovered from kefir (OG2; obtained from the ENA).  
245 Moreover, we found four isolates obtained from the ENA, two livestock-associated isolates from cows  
246 (BC05211 and NW19), one isolate recovered from kefir (OG2) and one isolate recovered from a willow  
247 plant (RIT283) were genetically related to ENA isolates which were recovered from the bloodstream.  
248 Interestingly, using the ENA, we were unable to find *S. haemolyticus* isolates that belonged to the same  
249 genetic lineage of isolate 506, which was recovered in public areas in the hospital in West London in  
250 this study.

251

252 PopPUNK analyses revealed that there were 38 combined core and accessory gene clusters predicted,  
 253 of which the accessory genome was found within 11 clusters (Figure 4). Five of these clusters included  
 254 isolates that were identified belonging to the same combined cluster and 6 clusters included isolates  
 255 that were only composed of those ENA isolates that have previously been recovered from clinical  
 256 samples (blood). Interestingly, isolates recovered from the East London Community and the ELH were  
 257 found together in the same cluster despite not always possessing the same combined core and accessory  
 258 cluster (cluster 3). Isolates recovered from West London were found in different clusters (clusters 2, 3  
 259 and 6). We observed genetic relatedness of the accessory genomes of all isolates recovered from East  
 260 London with those ENA isolates that have previously been recovered from an eye infection ( SH1572);  
 261 venous catheter (95671); and environmental isolates (a copper alloy coin; R1P1), whereas genetic  
 262 relatedness of the accessory genomes of isolates recovered from West London was observed with those  
 263 ENA isolates that were recovered from eye infections (SH1572 and SH1574), colon (1HT3),  
 264 bloodstream ( FDAARGOS-148), vagina (DNF00585), and sputum (C10F); healthy humans  
 265 (JCSC1435, MTCC 3383, SHN3) and plants (RIT283 and 167); livestock (NW19) and a companion  
 266 animal (SW007).

267

### 268 **Phylogenetic analysis of *S. hominis***

269 To determine the relatedness of *S. hominis* isolates in this study with those in the ENA, ten *mecA*<sup>+</sup> *S.*  
 270 *hominis* isolates recovered from public settings in East and West London in our study were  
 271 phylogenetically compared with *S. hominis* isolates recovered from different sources that have  
 272 previously been submitted to the ENA. This included isolates recovered from bloodstreams (n=11),  
 273 healthy human skin (n=6); livestock (cows; n=11), **kefir (n=1)** (mosquitos (n=3), natural environments  
 274 (ancient permafrost and an air sample from residential areas; n=2) and a plant isolate (rice seeds; n=1).  
 275 SNP core phylogenetic tree of *S. hominis* isolates contained 2 distinct clades: A and B (Figure 2). Clade  
 276 A consisted of isolates obtained from the ENA and previously recovered from livestock (cows), healthy  
 277 human skin, the air in residential areas, whereas clade B consisted of ENA isolates previously  
 278 recovered from clinical bloodstream infections, healthy human skin, **kefir** and rice seeds, mosquitos  
 279 and ancient permafrost. Among *mecA*<sup>+</sup> *S. hominis* isolates recovered in this study, only the isolate 385  
 280 from public areas in hospitals was found in clade A, whereas the remaining of the isolates were found  
 281 together in clade B. Isolates 387, 386, 620, 623 and 372 recovered from hospitals in East and West  
 282 London were found in the same subclade and were genetically related to ENA isolate that have been  
 283 previously recovered from healthy human skin (ZBW5). Isolates 207, 208 and 209 recovered from  
 284 West London public areas in the community have been found in the same subclade and were genetically  
 285 related to an isolate recovered from a skin of a healthy human (UMB022), environmental isolates  
 286 (ancient permafrost in Russia; MMP2), Asian Malaria Mosquito bodies (AS1, AS2 and AS3) and **kefir**  
 287 (KR) (Hughes et al., 2016; Kashuba et al., 2017). We did, however, observe that isolates 207, 208 and  
 288 209 in this subclade had divergence in their genetic relationship with the ENA isolates. Isolate 385  
 289 recovered from East London was genetically related to ENA isolates previously recovered from healthy  
 290 humans (Hudgins) and air samples from residential areas (H69). Interestingly, isolate 479 recovered  
 291 from public areas in a WLH was not genetically related to other isolates. All *S. hominis* ENA isolates  
 292 that were genetically related to isolates from public settings in London except the ENA isolate  
 293 recovered from mosquito and the permafrost have been shown to harbour the *mecA* gene. All *S. hominis*  
 294 ENA isolates that were previously recovered from clinical samples and 8 out of 10 ENA isolates  
 295 recovered from livestock (SNUC 2444, SNUC 5746, SNUC 3403, SNUC 5852, SNUC 4474, SNUC  
 296 2620, SNUC 5336 and SNUC 3870) were not genetically related to isolates that we recovered from  
 297 public settings or other ENA isolates we analysed.



298 PopPUNK analyses identified 23 combined core and accessory gene clusters, 5 of which were distinct  
299 clusters based on accessory genomes (Figure 6). Clinical isolates accessory genomes were found to be  
300 clustered together (cluster 5) distinct from those that were recovered from different sources.  
301 Additionally, 8 of the 11 *S. hominis* ENA isolates that have been previously recovered from livestock  
302 (SNUC 2444, SNUC 5746, SNUC 3403, SNUC 5852, SNUC 4474, SNUC 2620, SNUC 5336 and  
303 SNUC 3870) were clustered together (cluster 2). All but one (385) *mecA*<sup>+</sup> isolates recovered from  
304 public settings in hospitals in this study, were in the same accessory genome cluster (cluster 4). This  
305 cluster includes isolate 479 from WLH, which was shown not to be phylogenetically related to other  
306 isolates by its core genome. The isolates in cluster 4 in this study were recovered from public settings  
307 and were related to those ENA isolates that have previously been recovered from healthy human skin  
308 (ZBW5), rice seed (RE2.10) and air samples from residential areas (H69) by their accessory genome.

309

### 310 Accessory gene analysis of ENA of the *S. hominis* clinical isolates

311 Core genome phylogenetic analyses showed that *S. hominis* ENA isolates that were previously  
312 recovered from the hospital associated infections belonged to a single subclade and formed a single  
313 cluster by accessory genome analyses (PopPunk pipeline). Due to these observations, we decided to  
314 investigate these isolates further to identify whether they harboured any unique genes ubiquities to  
315 them/their accessory genome. We found 22 unique genes that were ubiquities to these *S. hominis*  
316 hospital associated infections ENA isolates. Fifteen of these genes were identified as being hypothetical  
317 (Supplementary information Table S3). Those that were identified as non-hypothetical genes included  
318 the chromosome recombinase gene *ccrA3/B3*; transposition regulatory protein allele *tnpB*; cadmium  
319 resistance genes *cadA*, *cadX* and *cadD* allele; putative DNA repair protein, *radC* and copper-sensing  
320 transcriptional repressor *ricR*.

321

### 322 Discussion

323 Genomic comparative studies are vital to aid our understanding of the relatedness of pathogenic  
324 bacteria recovered from different ecological niches and the transmission of these bacteria between  
325 human, livestock and the environment (Hartfield et al., 2014). To date, the majority of phylogenetic  
326 studies of CoNS has focused on studying bacteria recovered from clinical settings (Cavanagh et al.,  
327 2014a; Post et al., 2017). In addition, there is a limited number of studies reporting on community-  
328 associated and livestock-associated CoNS, but little is known about the genetic lineages of CoNS  
329 recovered from public settings (Conlan et al., 2012; Chaudhry and Patil, 2016). Using a comparative  
330 genome approach, we aimed to determine whether the *mecA*<sup>+</sup> CoNS recovered from public settings in  
331 East and West London in our study were genetically related to those isolates previously submitted to  
332 the ENA by others, including isolates recovered from different sources and those associated with  
333 infections (McEwen and Collignon, 2018).

334 In our study, we found that *S. epidermidis*, *S. haemolyticus* and *S. hominis* were the most common  
335 species of CoNS recovered from public settings that were *mecA* positive. This is consistent with reports  
336 that previously showed that these species were the most common isolates recovered from nosocomial  
337 infections and public settings (Xu et al., 2015; Asaad et al., 2016; Seng et al., 2017). Given that there  
338 is a limited number of WGS data available in the European Nucleotide Archive which has **the record**  
339 **of their isolation source** (<https://www.ebi.ac.uk/ena>) for *S. epidermidis*, *S. haemolyticus* and *S. hominis*  
340 isolated from different sources, we carried out phylogenetic analyses of these isolates only.

341

342 Core phylogenetic analysis of all three species revealed that *mecA* isolates recovered from these setting  
343 were genetically diverse and span across different clades. For *S. epidermidis* the majority of the isolates  
344 recovered from public settings in WLH (711, 712, 713, 715 and 716) were phylogenetically similar to  
345 ENA clinical isolates (bloodstream) by their core genome as well as their sequence types (ST2). The  
346 discovery of ST2 in public areas hospital areas is not surprising as it is the most common sequence  
347 types found in hospital-acquired infections (Deplano et al., 2016). However, it does suggest a route in  
348 which well-known hospital-acquired strains can reach the community [from a public area](#). We also  
349 found that *mecA*<sup>+</sup> isolates 435, 475, 631 recovered from the public areas in WLH were genetically  
350 related to clinical isolates recovered from bloodstream; isolates 436 recovered from West London was  
351 genetically related to isolate recovered from urine tract infection (FDAARGOS-83), whereas isolate  
352 465 was genetically related to an isolate recovered from endotracheal tube biofilm of a mechanically  
353 ventilated patient (ET-024). Additionally, ENA isolates Y24, PR246B0 M01 and M025 previously  
354 recovered from livestock and their housing was phylogenetically related to an isolate in this study that  
355 was recovered from public areas in hospitals (407). **These isolates belonged to the same sequence type**  
356 **(ST59), which has previously been associated with isolates recovered from both livestock and humans**  
357 **(Argudín et al., 2015b; Xu et al., 2018a). These findings indicate that isolates found in public areas in**  
358 **hospitals are genetically related to those that have previously been reported to be associated with**  
359 **infections in humans and livestock. Other studies have shown *mecA*<sup>+</sup> *S. epidermidis* as a common cause**  
360 **in bovine mastitis as well as has been recovered from cows milk (Feßler et al., 2010; Fernandes Dos**  
361 **Santos et al., 2016). In addition, pigs have also been shown to be a reservoir of *mecA*<sup>+</sup> *S. epidermidis***  
362 **that had similar virulence and antibiotic resistance gene profiles as isolates recovered from humans,**  
363 **largely indicating the transmission between humans and pigs (Tulinski et al., 2012; Argudín et al.,**  
364 **2015a). These reports, combined with our data, suggest that *S. epidermidis* may represent zoonoses**  
365 **and that livestock-associated *mecA*<sup>+</sup> *S. epidermidis* isolates belong to the same genetic lineages as the**  
366 **isolates that have been shown to cause infections in humans. Some known lineages that cause infections**  
367 **in humans may have originated in animals and have been transferred to humans and their associated**  
368 **environments either via direct contact of farmers with animals or via food.**

369  
370 For *S. haemolyticus* core genome phylogenetic analysis all but two isolates recovered from public  
371 settings in this study were genetically related to isolates recovered from an eye conjunctivitis  
372 (SH1572), bloodstream infection (M-176) and central venous catheter (95671) obtained from the ENA.  
373 These results show that the isolates recovered from public settings in this study may potentially pose a  
374 public health risk as they belong to the same genetic lineages that have been shown to cause eye  
375 conjunctivitis and bloodstream infections (Cavanagh et al., 2014b; Panda and Singh, 2016). This  
376 includes the 4 *mecA*<sup>+</sup> isolates that were recovered from general public settings in East London (1, 93,  
377 99 and 105), suggesting that isolates that cause bloodstream infections are not only present in public  
378 areas in hospitals but can also be found in general public settings. **Moreover, we also identified that**  
379 **isolates recovered from public areas in hospitals were genetically related to those isolates that have**  
380 **previously been recovered from a dog (SW007; ENA) and kefir, that as a fermented milk product**  
381 **commonly contains different species of CoNS (OG2; ENA), which have been shown to carry the *mecA***  
382 **gene (Prado et al., 2015). Previous reports have demonstrated that companion animals are potential**  
383 **reservoirs for the *mecA* gene, which can be transmitted to humans via contact as well as via food**  
384 **products (Ruzauskas et al., 2014). In this study, we were not able to determine whether those *S.***  
385 ***haemolyticus* isolates that were genetically related to an isolate recovered from a dog could pose a**  
386 **potential risk to public health as no previous studies have linked the genetic lineages of *S. haemolyticus***  
387 **isolates recovered from companion animals to that of isolates that have caused infections in humans.**  
388 **To the best of our knowledge, there are no reports that *S. haemolyticus* recovered from livestock**  
389 **belongs to the same genetic lineages known to cause infections in humans. In addition, we found that**

390 an isolate in the ENA recovered from kefir (OG2), was also related to the isolate in the ENA that has  
391 been recovered from clinical bloodstream infection (M-176).

392 Core genome phylogenetic analysis of *S. hominis* showed that the ENA livestock (cows) and ENA  
393 clinical isolates were genetically different to each other with none being related to the isolates  
394 recovered from public settings in East and West London suggesting that that the isolates from livestock  
395 and clinical samples have evolved separately and not crossed over into other niches. We did, however,  
396 find that our isolates from public setting to be phylogenetically related to isolates recovered from  
397 healthy humans skin (Hudgins and ZBW5), air samples from residential areas (H69), mosquitos'  
398 bodies (As1, As2 and As3), ancient permafrost (MMP2) and kefir (KR) (Hughes et al., 2016; Rivera-  
399 Perez et al., 2016). This suggests that mosquitoes could be possible vectors for transmitting *S. hominis*  
400 while feeding on their host and that genetically these *mecA*<sup>+</sup> *S. hominis* isolates recovered from general  
401 public settings have not evolved much since ancient times (Hughes et al., 2016). Mosquitos are vectors  
402 for viruses, protozoa and parasites that can spread and cause disease in humans and animals but  
403 currently, it is unreported if they can transfer and initiate bacterial infections (Torres-Guerrero et al.,  
404 2017; Assaid et al., 2020; Multini et al., 2020). The ENA isolates recovered from mosquitos (As1, As2  
405 and As3) were genetically related to an ENA isolate recovered from healthy human skin (UMB0272)  
406 (Figure 5). Therefore, we can conclude that the ENA isolates recovered from mosquitos belong to the  
407 same genetic lineages as those recovered from humans. In addition, the findings that the *mecA*<sup>+</sup> *S.*  
408 *hominis* isolates (385) in our study belonged to the same genetic lineage as the isolate recovered from  
409 an air sample in a residential area (H69) suggests that *S. hominis* can be transmitted through the air  
410 from humans to high-frequency touched surfaces or vice versa (Lymperopoulou et al., 2017).

411 Accessory genome is important for bacterial adaption and survival in different environments with  
412 studies of *Vibrio vulnificus*, *Legionella pneumophila* and *Pseudomonas aeruginosa* showing that clinical  
413 and environmental isolates can be distinguished by their accessory genomes (Kung et al., 2010; Koton et al.,  
414 2015; Mercante et al., 2018, 1). However, studies looking at accessory genomes of CoNS species recovered  
415 from different ecological niches are lacking. In the t-SNE plots, we observed that all the clusters generated in  
416 *S. epidermidis* analyses had a mixture of isolates from different sources, whereas *S. haemolyticus* and *S.*  
417 *hominis* analysis had clusters that were generated purely on isolates obtained from the ENA, which have been  
418 recovered from clinical bloodstream infection samples. In addition, there were two clusters of *S. hominis* ENA  
419 isolates that were only recovered from livestock (cows). This suggests that *S. epidermidis* isolates  
420 recovered from infections have previously been recovered from other niches. **This also coincides with**  
421 **a previously published study which showed that *S. epidermidis* infections are derived from a diverse**  
422 **genetic backgrounds that possess k-mers (infection-associated genetic elements) associated with**  
423 **pathogenicity traits (Méric et al., 2018).** These genes **have** likely originated from bacterial species  
424 associated with a particular niche and been transmitted to a new niche via humans or food. We did,  
425 however, observed clusters in the accessory genome that only contain *S. haemolyticus* and *S. hominis*  
426 ENA clinical associated isolates. This suggests that these isolates might have originated in nosocomial  
427 environments but have not spread to other non-hospital niches. This is further supported by the fact  
428 that *S. hominis* isolates in this study shared 22 unique genes with those isolates that have previously  
429 been recovered from different infections. We found that of these twenty-two genes, in particular those  
430 responsible for cadmium and copper resistance were the most interesting. Previous studies have shown  
431 the importance of cadmium resistance in *Helicobacter pylori* and *Listeria monocytogenes* virulence,  
432 whereas copper resistance is important for *S. aureus* survival within macrophages (Stähler et al., 2006;  
433 Purves et al., 2018). **Moreover, it has been shown that metal resistance genes (cadmium, arsenic and**  
434 **zinc) are extensively exchanged between clinical associated *S. epidermidis* and *S. aureus* isolates and**  
435 **may play a role in their survival within hospital settings (Méric et al., 2015).** However, it is currently  
436 **unknown whether these genes contribute to the virulence or survival of *S. hominis* or *S. haemolyticus***  
437 **within clinical settings.** We found that *S. haemolyticus* isolates recovered from public settings in East

438 London occupied the same cluster, whereas all but one *S. hominis* isolate from public areas in hospitals  
439 were found within the same cluster. This indicates that these species in these areas possess a similar  
440 pool of genes that are horizontally transferred due to the similarities in the microbiome in that  
441 geographical area or environment and/or adaptation required to survive in these niches (Segerman,  
442 2012). In addition, we found that *S. epidermidis* and *S. haemolyticus* recovered from public settings in  
443 this study had similar accessory genomes with those isolates that have previously been recovered from  
444 different infections (*S. epidermidis*: bloodstream, urinary tract infections and cerebrospinal fluid; *S.*  
445 *haemolyticus*: bloodstream and eye infections) healthy humans, animals (*S. epidermidis* and *S.*  
446 *haemolyticus*: livestock; *S. haemolyticus*: companion animals), plants and wider environments. Both  
447 core and accessory genomes phylogenetic analyses suggested that *mecA*<sup>+</sup> isolates in this study  
448 originated from different ecological niches. Therefore, we hypothesis that these isolates contain genes  
449 associated with bacterial species across many different genera from different environments. These  
450 genes can horizontally be transferred to other CoNS isolates in public settings.

451 This study implies that CoNS bacteria from high frequency touched surfaces may be a potential risk to  
452 public health. **Although, they lack virulence determinants, many cases of CoNS infections have been**  
453 **reported inside and outside of hospital settings (Rogers et al., 2009).** In addition, CoNS harbour many  
454 types of antibiotic resistance genes which can horizontally be transferred across to more virulent  
455 pathogens (Xu et al., 2018b; Lee et al., 2019). It also plausible that the CoNS strains we recovered  
456 from public settings may have originated from animals either from livestock or companion animals.  
457 Animals can be a reservoir of antibiotic-resistant CoNS isolates which can be transmitted to humans  
458 furthering the spread of antibiotic-resistance bacteria (Bhargava and Zhang, 2012; Argudín et al.,  
459 2015a). **The results of this study can be used to review the hand hygiene practices adapted by the**  
460 **general public; the design of the build environments to reduce infections and the use of appropriate**  
461 **disinfectants to prevent further transmission of potentially pathogenic bacteria in these areas.**

### 462 **Conclusion**

463 *MecA*<sup>+</sup> *S. epidermidis* and *S. haemolyticus* isolates recovered from public settings in the community  
464 and hospitals in this study may pose a potential health risk as we showed that they belong to the same  
465 genetic lineages of those isolates that have previously been recovered from different infections (data  
466 obtained from the ENA). However, in this study we did not find similar features for the *S. hominis*  
467 *mecA*<sup>+</sup> isolates recovered from public settings. In addition, we showed the relatedness of the *mecA*<sup>+</sup>  
468 staphylococcal isolates (all three species) in this study with isolates recovered from livestock and  
469 healthy humans (ENA). Further studies are warranted to aid our understanding of whether the isolates  
470 recovered in this study that were genetically related to those recovered from livestock (ENA) could  
471 potentially cause infections in humans. The results from this study should be used to instigate a review  
472 on how the public wash their hands, the design of public areas (including ventilation practices) and  
473 how disinfectants are used to improve the **hygiene practices** in these areas.

### 474 **Acknowledgment**

475 Rory Cave is funded by the UEL PhD studentship awarded to HVM.

### 476 **Reference**

477 Abu-Saleh, R., Nitzan, O., Saliba, W., Colodner, R., Keness, Y., Yanovskay, A., et al. (2018).  
478 Bloodstream Infections Caused by Contaminants: Epidemiology and Risk Factors: A 10-Year  
479 Surveillance. *Isr. Med. Assoc. J.* 20, 433–437.

- 480 Altschul, S. F., Gish, W., Miller, W., Myers, E. W., and Lipman, D. J. (1990). Basic local alignment  
481 search tool. *J. Mol. Biol.* 215, 403–410. doi:10.1016/S0022-2836(05)80360-2.
- 482 Andrews, S. (2011). *FastQC A Quality Control tool for High Throughput Sequence Data*.  
483 Cambridge, UK: Babraham Institute Available at:  
484 <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/> [Accessed September 23, 2019].
- 485 Argimón, S., Abudahab, K., Goater, R. J. E., Fedosejev, A., Bhai, J., Glasner, C., et al. (2016).  
486 Microreact: visualizing and sharing data for genomic epidemiology and phylogeography.  
487 *Microb Genom* 2. doi:10.1099/mgen.0.000093.
- 488 Argudín, M. A., Vanderhaeghen, W., and Butaye, P. (2015a). Antimicrobial resistance and  
489 population structure of *Staphylococcus epidermidis* recovered from pig farms in Belgium.  
490 *Vet. J.* 203, 302–308. doi:10.1016/j.tvjl.2015.01.008.
- 491 Argudín, M. A., Vanderhaeghen, W., Vandendriessche, S., Vandecandelaere, I., André, F.-X., Denis,  
492 O., et al. (2015b). Antimicrobial resistance and population structure of *Staphylococcus*  
493 *epidermidis* recovered from animals and humans. *Vet. Microbiol.* 178, 105–113.  
494 doi:10.1016/j.vetmic.2015.04.019.
- 495 Asaad, A. M., Ansar Qureshi, M., and Mujeeb Hasan, S. (2016). Clinical significance of coagulase-  
496 negative staphylococci isolates from nosocomial bloodstream infections. *Infect Dis (Lond)* 48,  
497 356–360. doi:10.3109/23744235.2015.1122833.
- 498 Assaid, N., Mousson, L., Moutailler, S., Arich, S., Akarid, K., Monier, M., et al. (2020). Evidence of  
499 circulation of West Nile virus in *Culex pipiens* mosquitoes and horses in Morocco. *Acta*  
500 *Trop.*, 105414. doi:10.1016/j.actatropica.2020.105414.
- 501 Bankevich, A., Nurk, S., Antipov, D., Gurevich, A. A., Dvorkin, M., Kulikov, A. S., et al. (2012).  
502 SPAdes: A New Genome Assembly Algorithm and Its Applications to Single-Cell  
503 Sequencing. *J Comput Biol* 19, 455–477. doi:10.1089/cmb.2012.0021.
- 504 Becker, K., Heilmann, C., and Peters, G. (2014). Coagulase-Negative Staphylococci. *Clinical*  
505 *Microbiology Reviews* 27, 870–926. doi:10.1128/CMR.00109-13.
- 506 Bhargava, K., and Zhang, Y. (2012). Multidrug-resistant coagulase-negative Staphylococci in food  
507 animals. *Journal of Applied Microbiology* 113, 1027–1036. doi:10.1111/j.1365-  
508 2672.2012.05410.x.
- 509 Bolger, A. M., Lohse, M., and Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina  
510 sequence data. *Bioinformatics* 30, 2114–2120. doi:10.1093/bioinformatics/btu170.
- 511 Cavanagh, J. P., Hjerde, E., Holden, M. T. G., Kahlke, T., Klingenberg, C., Flægstad, T., et al.  
512 (2014a). Whole-genome sequencing reveals clonal expansion of multiresistant  
513 *Staphylococcus haemolyticus* in European hospitals. *J Antimicrob Chemother* 69, 2920–2927.  
514 doi:10.1093/jac/dku271.
- 515 Cavanagh, J. P., Hjerde, E., Holden, M. T., Kahlke, T., Klingenberg, C., Flægstad, T., et al. (2014b).  
516 Whole-genome sequencing reveals clonal expansion of multiresistant *Staphylococcus*

- 517 *haemolyticus* in European hospitals. *J Antimicrob Chemother* 69, 2920–2927.  
518 doi:10.1093/jac/dku271.
- 519 Cave, R., Misra, R., Chen, J., Wang, S., and Mkrtychyan, H. V. (2019). Whole genome sequencing  
520 revealed new molecular characteristics in multidrug resistant staphylococci recovered from  
521 high frequency touched surfaces in London. *Sci Rep* 9, 1–13. doi:10.1038/s41598-019-45886-  
522 6.
- 523 Chaudhry, V., and Patil, P. B. (2016). Genomic investigation reveals evolution and lifestyle  
524 adaptation of endophytic *Staphylococcus epidermidis*. *Scientific Reports* 6, 19263.  
525 doi:10.1038/srep19263.
- 526 Chaves, F., Garcia-Alvarez, M., Sanz, F., Alba, C., and Otero, J. R. (2005). Nosocomial Spread of a  
527 *Staphylococcus hominis* subsp. novobiosepticus Strain Causing Sepsis in a Neonatal Intensive  
528 Care Unit. *Journal of Clinical Microbiology* 43, 4877–4879. doi:10.1128/JCM.43.9.4877-  
529 4879.2005.
- 530 Cherifi, S., Byl, B., Deplano, A., Nonhoff, C., Denis, O., and Hallin, M. (2013). Comparative  
531 Epidemiology of *Staphylococcus epidermidis* Isolates from Patients with Catheter-Related  
532 Bacteremia and from Healthy Volunteers. *Journal of Clinical Microbiology* 51, 1541–1547.  
533 doi:10.1128/JCM.03378-12.
- 534 Conlan, S., Mijares, L. A., Becker, J., Blakesley, R. W., Bouffard, G. G., Brooks, S., et al. (2012).  
535 *Staphylococcus epidermidis* pan-genome sequence analysis reveals diversity of skin  
536 commensal and hospital infection-associated isolates. *Genome Biol* 13, R64. doi:10.1186/gb-  
537 2012-13-7-r64.
- 538 Croucher, N. J., Page, A. J., Connor, T. R., Delaney, A. J., Keane, J. A., Bentley, S. D., et al. (2015).  
539 Rapid phylogenetic analysis of large samples of recombinant bacterial whole genome  
540 sequences using Gubbins. *Nucleic Acids Res.* 43, e15. doi:10.1093/nar/gku1196.
- 541 Cui, J., Liang, Z., Mo, Z., and Zhang, J. (2019). The species distribution, antimicrobial resistance and  
542 risk factors for poor outcome of coagulase-negative staphylococci bacteraemia in China.  
543 *Antimicrobial Resistance & Infection Control* 8, 65. doi:10.1186/s13756-019-0523-5.
- 544 Czekaj, T., Ciszewski, M., and Szewczyk, E. M. (2015). *Staphylococcus haemolyticus* – an emerging  
545 threat in the twilight of the antibiotics age. *Microbiology* 161, 2061–2068.  
546 doi:10.1099/mic.0.000178.
- 547 Deplano, A., Vandendriessche, S., Nonhoff, C., Dodémont, M., Roisin, S., and Denis, O. (2016).  
548 National surveillance of *Staphylococcus epidermidis* recovered from bloodstream infections  
549 in Belgian hospitals. *J. Antimicrob. Chemother.* 71, 1815–1819. doi:10.1093/jac/dkw086.
- 550 English Surveillance Programme for Antimicrobial Utilisation and Resistance (ESPAUR) (2019).  
551 Public Health England Wellington House 133-155 Waterloo Road London SE1 8UG: Public  
552 Health England Available at:  
553 [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/  
554 file/843129/English\\_Surveillance\\_Programme\\_for\\_Antimicrobial\\_Utilisation\\_and\\_Resistanc  
555 e\\_2019.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/843129/English_Surveillance_Programme_for_Antimicrobial_Utilisation_and_Resistance_2019.pdf).

- 556 Fernandes Dos Santos, F., Mendonça, L. C., Reis, D. R. de L., Guimarães, A. de S., Lange, C. C.,  
 557 Ribeiro, J. B., et al. (2016). Presence of mecA-positive multidrug-resistant *Staphylococcus*  
 558 *epidermidis* in bovine milk samples in Brazil. *J. Dairy Sci.* 99, 1374–1382.  
 559 doi:10.3168/jds.2015-9931.
- 560 Feßler, A. T., Billerbeck, C., Kadlec, K., and Schwarz, S. (2010). Identification and characterization  
 561 of methicillin-resistant coagulase-negative staphylococci from bovine mastitis. *J Antimicrob*  
 562 *Chemother* 65, 1576–1582. doi:10.1093/jac/dkq172.
- 563 Fitzgerald, J. R. (2012). Livestock-associated *Staphylococcus aureus*: origin, evolution and public  
 564 health threat. *Trends in Microbiology* 20, 192–198. doi:10.1016/j.tim.2012.01.006.
- 565 Gurevich, A., Saveliev, V., Vyahhi, N., and Tesler, G. (2013). QUAST: quality assessment tool for  
 566 genome assemblies. *Bioinformatics* 29, 1072–1075. doi:10.1093/bioinformatics/btt086.
- 567 Hanssen, A.-M., Kjeldsen, G., and Sollid, J. U. E. (2004). Local variants of Staphylococcal cassette  
 568 chromosome mec in sporadic methicillin-resistant *Staphylococcus aureus* and methicillin-  
 569 resistant coagulase-negative Staphylococci: evidence of horizontal gene transfer? *Antimicrob.*  
 570 *Agents Chemother.* 48, 285–296. doi:10.1128/aac.48.1.285-296.2004.
- 571 Hartfield, M., Murall, C. L., and Alizon, S. (2014). Clinical applications of pathogen phylogenies.  
 572 *Trends in Molecular Medicine* 20, 394–404. doi:10.1016/j.molmed.2014.04.002.
- 573 Hughes, G. L., Raygoza, J. G., Koundal, V., Rasgon, J. L., and Mwangi, M. M. (2016). Genome  
 574 Sequences of *Staphylococcus hominis* Strains ShAs1, ShAs2, and ShAs3, Isolated from the  
 575 Asian Malaria Mosquito *Anopheles stephensi*. *Genome Announc* 4.  
 576 doi:10.1128/genomeA.00085-16.
- 577 Janda, J. M., and Abbott, S. L. (2007). 16S rRNA Gene Sequencing for Bacterial Identification in the  
 578 Diagnostic Laboratory: Pluses, Perils, and Pitfalls. *Journal of Clinical Microbiology* 45,  
 579 2761–2764. doi:10.1128/JCM.01228-07.
- 580 Kashuba, E., Dmitriev, A. A., Kamal, S. M., Melefors, O., Griva, G., Römling, U., et al. (2017).  
 581 Ancient permafrost staphylococci carry antibiotic resistance genes. *Microb Ecol Health Dis*  
 582 28. doi:10.1080/16512235.2017.1345574.
- 583 Koboldt, D. C., Chen, K., Wylie, T., Larson, D. E., McLellan, M. D., Mardis, E. R., et al. (2009).  
 584 VarScan: variant detection in massively parallel sequencing of individual and pooled samples.  
 585 *Bioinformatics* 25, 2283–2285. doi:10.1093/bioinformatics/btp373.
- 586 Koton, Y., Gordon, M., Chalifa-Caspi, V., and Bisharat, N. (2015). Comparative genomic analysis of  
 587 clinical and environmental *Vibrio vulnificus* isolates revealed biotype 3 evolutionary  
 588 relationships. *Front. Microbiol.* 5. doi:10.3389/fmicb.2014.00803.
- 589 Kung, V. L., Ozer, E. A., and Hauser, A. R. (2010). The Accessory Genome of *Pseudomonas*  
 590 *aeruginosa*. *Microbiol. Mol. Biol. Rev.* 74, 621–641. doi:10.1128/MMBR.00027-10.
- 591 Land Area and Population Density, Ward and Borough - London Datastore Available at:  
 592 <https://data.london.gov.uk/dataset/land-area-and-population-density-ward-and-borough>  
 593 [Accessed May 11, 2020].

- 594 Lee, J. Y. H., Monk, I. R., Gonçalves da Silva, A., Seemann, T., Chua, K. Y. L., Kearns, A., et al.  
595 (2018). Global spread of three multidrug-resistant lineages of *Staphylococcus epidermidis*.  
596 *Nat Microbiol* 3, 1175–1185. doi:10.1038/s41564-018-0230-7.
- 597 Lee, J.-H., Heo, S., Jeong, M., and Jeong, D.-W. (2019). Transfer of a mobile *Staphylococcus*  
598 *saprophyticus* plasmid isolated from fermented seafood that confers tetracycline resistance.  
599 *PLOS ONE* 14, e0213289. doi:10.1371/journal.pone.0213289.
- 600 Lees, J. A., Harris, S. R., Tonkin-Hill, G., Gladstone, R. A., Lo, S. W., Weiser, J. N., et al. (2019).  
601 Fast and flexible bacterial genomic epidemiology with PopPUNK. *Genome Res.* 29, 304–316.  
602 doi:10.1101/gr.241455.118.
- 603 Lei, H., Li, Y., Xiao, S., Yang, X., Lin, C., Norris, S. L., et al. (2017). Logistic growth of a surface  
604 contamination network and its role in disease spread. *Scientific Reports* 7, 1–10.  
605 doi:10.1038/s41598-017-13840-z.
- 606 Letunic, I., and Bork, P. (2016). Interactive tree of life (iTOL) v3: an online tool for the display and  
607 annotation of phylogenetic and other trees. *Nucleic Acids Res.* 44, W242-245.  
608 doi:10.1093/nar/gkw290.
- 609 Lymperopoulou, D. S., Coil, D. A., Schichnes, D., Lindow, S. E., Jospin, G., Eisen, J. A., et al.  
610 (2017). Draft genome sequences of eight bacteria isolated from the indoor environment:  
611 *Staphylococcus capitis* strain H36, *S. capitis* strain H65, *S. cohnii* strain H62, *S. hominis*  
612 strain H69, *Microbacterium* sp. strain H83, *Mycobacterium iranicum* strain H39, *Plantibacter*  
613 sp. strain H53, and *Pseudomonas oryzihabitans* strain H72. *Stand Genomic Sci* 12.  
614 doi:10.1186/s40793-017-0223-9.
- 615 McEwen, S. A., and Collignon, P. J. (2018). Antimicrobial Resistance: a One Health Perspective.  
616 *Microbiol Spectr* 6. doi:10.1128/microbiolspec.ARBA-0009-2017.
- 617 Mercante, J. W., Caravas, J. A., Ishaq, M. K., Kozak-Muiznieks, N. A., Raphael, B. H., and  
618 Winchell, J. M. (2018). Genomic heterogeneity differentiates clinical and environmental  
619 subgroups of *Legionella pneumophila* sequence type 1. *PLOS ONE* 13, e0206110.  
620 doi:10.1371/journal.pone.0206110.
- 621 Méric, G., Mageiros, L., Pensar, J., Laabei, M., Yahara, K., Pascoe, B., et al. (2018). Disease-  
622 associated genotypes of the commensal skin bacterium *Staphylococcus epidermidis*. *Nat*  
623 *Commun* 9. doi:10.1038/s41467-018-07368-7.
- 624 Méric, G., Miragaia, M., de Been, M., Yahara, K., Pascoe, B., Mageiros, L., et al. (2015). Ecological  
625 Overlap and Horizontal Gene Transfer in *Staphylococcus aureus* and *Staphylococcus*  
626 *epidermidis*. *Genome Biol Evol* 7, 1313–1328. doi:10.1093/gbe/evv066.
- 627 Multini, L. C., Wilke, A. B. B., and Marrelli, M. T. (2020). Neotropical Anopheles (Kerteszia)  
628 mosquitoes associated with bromeliad-malaria transmission in a changing world. *Acta Trop.*,  
629 105413. doi:10.1016/j.actatropica.2020.105413.
- 630 Nanoukon, C., Argemi, X., Sogbo, F., Orekan, J., Keller, D., Affolabi, D., et al. (2017). Pathogenic  
631 features of clinically significant coagulase-negative staphylococci in hospital and community



- 632 infections in Benin. *International Journal of Medical Microbiology* 307, 75–82.  
633 doi:10.1016/j.ijmm.2016.11.001.
- 634 Otto, M. (2009). *Staphylococcus epidermidis* – the “accidental” pathogen. *Nat Rev Microbiol* 7, 555–  
635 567. doi:10.1038/nrmicro2182.
- 636 Page, A. J., Cummins, C. A., Hunt, M., Wong, V. K., Reuter, S., Holden, M. T. G., et al. (2015).  
637 Roary: rapid large-scale prokaryote pan genome analysis. *Bioinformatics* 31, 3691–3693.  
638 doi:10.1093/bioinformatics/btv421.
- 639 Pain, M., Hjerde, E., Klingenberg, C., and Cavanagh, J. P. (2019). Comparative Genomic Analysis of  
640 *Staphylococcus haemolyticus* Reveals Key to Hospital Adaptation and Pathogenicity. *Front.*  
641 *Microbiol.* 10. doi:10.3389/fmicb.2019.02096.
- 642 Panda, S., and Singh, D. V. (2016). Whole-Genome Sequences of *Staphylococcus haemolyticus*  
643 Isolated from Infected Eyes and Healthy Conjunctiva in Bhubaneswar, India. *Genome*  
644 *Announc* 4. doi:10.1128/genomeA.00099-16.
- 645 Post, V., Harris, L. G., Morgenstern, M., Mageiros, L., Hitchings, M. D., Méric, G., et al. (2017).  
646 Comparative Genomics Study of *Staphylococcus epidermidis* Isolates from Orthopedic-  
647 Device-Related Infections Correlated with Patient Outcome. *Journal of Clinical Microbiology*  
648 55, 3089–3103. doi:10.1128/JCM.00881-17.
- 649 Prado, M. R., Blandón, L. M., Vandenberghe, L. P. S., Rodrigues, C., Castro, G. R., Thomaz-Soccol,  
650 V., et al. (2015). Milk kefir: composition, microbial cultures, biological activities, and related  
651 products. *Front. Microbiol.* 6. doi:10.3389/fmicb.2015.01177.
- 652 Purves, J., Thomas, J., Riboldi, G. P., Zapotoczna, M., Tarrant, E., Andrew, P. W., et al. (2018). A  
653 horizontally gene transferred copper resistance locus confers hyper-resistance to antibacterial  
654 copper toxicity and enables survival of community acquired methicillin resistant  
655 *Staphylococcus aureus* USA300 in macrophages. *Environ Microbiol* 20, 1576–1589.  
656 doi:10.1111/1462-2920.14088.
- 657 Quainoo, S., Coolen, J. P. M., Hijum, S. A. F. T. van, Huynen, M. A., Melchers, W. J. G., Schaik, W.  
658 van, et al. (2017). Whole-Genome Sequencing of Bacterial Pathogens: the Future of  
659 Nosocomial Outbreak Analysis. *Clinical Microbiology Reviews* 30, 1015–1063.  
660 doi:10.1128/CMR.00016-17.
- 661 RIVERA-PEREZ, J. I., SANTIAGO-RODRIGUEZ, T. M., and TORANZOS, G. A. (2016).  
662 Paleomicrobiology: a Snapshot of Ancient Microbes and Approaches to Forensic  
663 Microbiology. *Microbiol Spectr* 4. doi:10.1128/microbiolspec.EMF-0006-2015.
- 664 Rogers, K. L., Fey, P. D., and Rupp, M. E. (2009). Coagulase-Negative Staphylococcal Infections.  
665 *Infectious Disease Clinics of North America* 23, 73–98. doi:10.1016/j.idc.2008.10.001.
- 666 Ruzauskas, M., Siugzdiniene, R., Klimiene, I., Virgailis, M., Mockeliunas, R., Vaskeviciute, L., et al.  
667 (2014). Prevalence of methicillin-resistant *Staphylococcus haemolyticus* in companion  
668 animals: a cross-sectional study. *Ann. Clin. Microbiol. Antimicrob.* 13, 56.  
669 doi:10.1186/s12941-014-0056-y.

- 670 Schoenfelder, S. M. K., Dong, Y., Feßler, A. T., Schwarz, S., Schoen, C., Köck, R., et al. (2017).  
 671 Antibiotic resistance profiles of coagulase-negative staphylococci in livestock environments.  
 672 *Vet. Microbiol.* 200, 79–87. doi:10.1016/j.vetmic.2016.04.019.
- 673 Segerman, B. (2012). The genetic integrity of bacterial species: the core genome and the accessory  
 674 genome, two different stories. *Front Cell Infect Microbiol* 2. doi:10.3389/fcimb.2012.00116.
- 675 Seng, R., Leungtongkam, U., Thummeepak, R., Chatdumrong, W., and Sitthisak, S. (2017). High  
 676 prevalence of methicillin-resistant coagulase-negative staphylococci isolated from a  
 677 university environment in Thailand. *Int. Microbiol.* 20, 65–73. doi:10.2436/20.1501.01.286.
- 678 Stähler, F. N., Odenbreit, S., Haas, R., Wilrich, J., Vliet, A. H. M. V., Kusters, J. G., et al. (2006).  
 679 The Novel *Helicobacter pylori* CznABC Metal Efflux Pump Is Required for Cadmium, Zinc,  
 680 and Nickel Resistance, Urease Modulation, and Gastric Colonization. *Infect Immun* 74, 3845–  
 681 3852. doi:10.1128/IAI.02025-05.
- 682 Stamatakis, A. (2014). RAxML version 8: a tool for phylogenetic analysis and post-analysis of large  
 683 phylogenies. *Bioinformatics* 30, 1312–1313. doi:10.1093/bioinformatics/btu033.
- 684 Torres-Guerrero, E., Quintanilla-Cedillo, M. R., Ruiz-Esmenjaud, J., and Arenas, R. (2017).  
 685 Leishmaniasis: a review. *F1000Res* 6. doi:10.12688/f1000research.11120.1.
- 686 Tulinski, P., Fluit, A. C., Wagenaar, J. A., Mevius, D., Vijver, L. van de, and Duim, B. (2012).  
 687 Methicillin-Resistant Coagulase-Negative Staphylococci on Pig Farms as a Reservoir of  
 688 Heterogeneous Staphylococcal Cassette Chromosome mec Elements. *Appl. Environ.*  
 689 *Microbiol.* 78, 299–304. doi:10.1128/AEM.05594-11.
- 690 Xu, Z., Misra, R., Jamrozy, D., Paterson, G. K., Cutler, R. R., Holmes, M. A., et al. (2018a). Whole  
 691 Genome Sequence and Comparative Genomics Analysis of Multi-drug Resistant  
 692 Environmental *Staphylococcus epidermidis* ST59. *G3: Genes, Genomes, Genetics* 8, 2225–  
 693 2230. doi:10.1534/g3.118.200314.
- 694 Xu, Z., Mkrtychyan, H. V., and Cutler, R. R. (2015). Antibiotic resistance and *mecA* characterization  
 695 of coagulase-negative staphylococci isolated from three hotels in London, UK. *Front*  
 696 *Microbiol* 6. doi:10.3389/fmicb.2015.00947.
- 697 Xu, Z., Shah, H. N., Misra, R., Chen, J., Zhang, W., Liu, Y., et al. (2018b). The prevalence, antibiotic  
 698 resistance and *mecA* characterization of coagulase negative staphylococci recovered from  
 699 non-healthcare settings in London, UK. *Antimicrobial Resistance & Infection Control* 7, 73.  
 700 doi:10.1186/s13756-018-0367-4.

701 **Tables**

702

703

704

705 **Table 1: *mecA*<sup>+</sup> coagulase negative staphylococcal isolates recovered from East and West**  
 706 **London public settings**

Isolate ID	Species ( <i>mecA</i> <sup>+</sup> )	Public settings in London
1	<i>S. haemolyticus</i>	ELC
93	<i>S. haemolyticus</i>	ELC
99	<i>S. haemolyticus</i>	ELC
105	<i>S. haemolyticus</i>	ELC
207	<i>S. hominis</i>	WLC
208	<i>S. hominis</i>	WLC
209	<i>S. hominis</i>	WLC
211	<i>S. cohnii</i>	WLC
321	<i>S. epidermidis</i>	ELH
327	<i>S. epidermidis</i>	ELH
329	<i>S. epidermidis</i>	ELH
343	<i>S. cohnii</i>	ELH
349	<i>S. cohnii</i>	ELH
355	<i>S. epidermidis</i>	ELH
361	<i>S. haemolyticus</i>	ELH
372	<i>S. hominis</i>	ELH
373	<i>S. haemolyticus</i>	ELH
385	<i>S. hominis</i>	ELH
386	<i>S. hominis</i>	ELH
387	<i>S. hominis</i>	ELH
407	<i>S. epidermidis</i>	ELH

## Phylogenetic study of staphylococci from public settings

435	<i>S. epidermidis</i>	WLH
436	<i>S. epidermidis</i>	WLH
445	<i>S. haemolyticus</i>	WLH
465	<i>S. epidermidis</i>	WLH
475	<i>S. epidermidis</i>	WLH
479	<i>S. hominis</i>	WLH
492	<i>S. haemolyticus</i>	WLH
506	<i>S. haemolyticus</i>	WLH
538	<i>S. haemolyticus</i>	WLH
620	<i>S. hominis</i>	WLH
623	<i>S. hominis</i>	WLH
631	<i>S. epidermidis</i>	WLH
664	<i>S. epidermidis</i>	WLH
673	<i>S. epidermidis</i>	WLH
699	<i>S. warneri</i>	WLH
700	<i>S. warneri</i>	WLH
702	<i>S. warneri</i>	WLH
711	<i>S. epidermidis</i>	WLH
712	<i>S. epidermidis</i>	WLH
713	<i>S. epidermidis</i>	WLH
715	<i>S. epidermidis</i>	WLH
716	<i>S. epidermidis</i>	WLH

707 ELC= East London Community; WLC= West London Community ELH= East London Hospital;

708 WLH=West London Hospital

