

UWL REPOSITORY
repository.uwl.ac.uk

Conjugative transfer frequencies of *mef(A)*-containing Tn1207.3 to macrolide-susceptible *Streptococcus pyogenes* belonging to different emm types

Hadjirin, Nazreen F., Harrison, E.M., Holmes, M.A. and Paterson, G.K. (2014) Conjugative transfer frequencies of *mef(A)*-containing Tn1207.3 to macrolide-susceptible *Streptococcus pyogenes* belonging to different emm types. *Letters in Applied Microbiology*, 58 (4). pp. 299-302. ISSN 0266-8254

<http://dx.doi.org/10.1111/lam.12213>

This is the Published Version of the final output.

UWL repository link: <https://repository.uwl.ac.uk/id/eprint/3027/>

Alternative formats: If you require this document in an alternative format, please contact: open.research@uwl.ac.uk

Copyright: Creative Commons: Attribution 4.0

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy: If you believe that this document breaches copyright, please contact us at open.research@uwl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Rights Retention Statement:

ORIGINAL ARTICLE

Conjugative transfer frequencies of *mef(A)*-containing Tn1207.3 to macrolide-susceptible *Streptococcus pyogenes* belonging to different *emm* types

N.F. Hadjirin¹, E.M. Harrison², M.A. Holmes² and G.K. Paterson²

¹ University of West London, London, UK

² Department of Veterinary Medicine, University of Cambridge, Cambridge, UK

Significance and Impact of the Study: The spread of antimicrobial resistance among pathogenic bacteria is an important problem, but the mechanisms of horizontal transfer between strains and species are often poorly understood. For instance, little is known on how macrolide resistance spreads between strains of the human pathogen *Strep. pyogenes* and why certain strains more commonly display resistance than others. Here, we show that *Strep. pyogenes* strains vary greatly in their ability to acquire a transposon encoding macrolide resistance by horizontal gene transfer *in vitro*. These data provide a novel insight into the transfer of antibiotic resistance between bacterial strains and offer an explanation for the differences in the frequency of resistance determinates and resistance seen among clinical isolates.

Keywords

antibiotics, molecular epidemiology, streptococci, transposons.

Correspondence

Nazreen F. Hadjirin and Gavin K. Paterson, Department of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge CB3 0ES, UK.

E-mails: nh396@cam.ac.uk and gkp27@cam.ac.uk

Present address

N.F. Hadjirin, Department of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge, CB3 0ES, UK

2013/2173: received 29 October 2013, revised 9 December 2013 and accepted 23 December 2013

doi:10.1111/lam.12213

Introduction

Streptococcus pyogenes (group A streptococci/GAS) remains an important human pathogen responsible for a wide variety of invasive and noninvasive infections including pharyngotonsillitis, skin and soft tissue infections and bacteraemia. Macrolides are widely used to treat GAS infections in patients allergic to β -lactam antibiotics.

Abstract

The aim of this study was to examine the gene transfer potential of *mef(A)*-containing Tn120.3 to macrolide-susceptible *Streptococcus pyogenes* belonging to different *emm* types. Using the filter mating technique, Tn1207.3 was transferred by conjugation to 23 macrolide-susceptible recipients representing 11 *emm* types. PCR analysis confirmed the presence of the *mef(A)* gene and the *comEC* junction regions of the Tn1207.3 insertion in resultant transconjugants. Significant variation was found in the transfer frequency of Tn1207.3 to different *Strep. pyogenes* strains, and this phenomenon may contribute to the differences in *mef(A)* frequency observed among clinical isolates.

The development and spread of macrolide resistance among GAS throughout the world is therefore a major concern.

The drug efflux pump, encoded by the *mef* gene (currently represented by four subclasses A, E, I and O), is one of the most common mechanisms responsible for resistance to 14- and 15-membered macrolides in GAS and other species (Sutcliffe *et al.* 1996a; Varaldo *et al.*

2009). At least three mobile elements carrying the *mef(A)* gene have been identified in GAS: the Tn1207.3 transposon (Santagati *et al.* 2003), the Φ 10394.4 phage chimera (Banks *et al.* 2003) and the Φ m46.1 element (Brenciani *et al.* 2010). Studies indicate a significant prevalence of the *mef(A)* gene among GAS isolates in Europe and elsewhere. For instance, nationwide surveillance in Germany noted a frequency of 31% (Bley *et al.* 2011), while an Italian study found the prevalence of *mef(A)* among GAS to be 16% (Creti *et al.* 2005). Interestingly, there appears to be a skewed distribution of macrolide resistance genes among different GAS strains belonging to different *emm* types. The *mef(A)* gene is most frequently harboured by certain *emm* types such as *emm1*, *emm2*, *emm3*, *emm4*, *emm9*, *emm12* and *emm75*, whereas *emm* types such as *emm22*, *emm77*, *emm87* and *emm89* have not yet been documented to carry any *mef* determinant (Creti *et al.* 2005; Grivea *et al.* 2006; Silva-Costa *et al.* 2008; Wajima *et al.* 2013). These observations suggest differences in the capacity of GAS *emm* types to acquire *mef* genes, and while the horizontal gene transfer of *mef(A)*-bearing genetic elements has been demonstrated *in vitro* by conjugation (Giovanetti *et al.* 2003; Santagati *et al.* 2003) and by transduction (Di Luca *et al.* 2010), no comparison has yet been made of *mef* gene transfer frequencies among different GAS strains.

The aim of this study therefore was to examine the transfer frequency of *mef(A)*-containing Tn1207.3 to diverse macrolide-susceptible recipients representing a variety of clinically important *emm* types.

Results and discussion

Erythromycin-resistant transconjugants were detected in 21/23 of the tested recipient strains, with mean frequencies of transconjugants ranging from 7.20×10^{-8} to 1.13×10^{-6} conjugant/recipient (Table 1). Statistical analysis showed that there was a significant difference in the conjugation frequency between strains (ANOVA, $P < 0.001$). PCR confirmed that all 85 randomly selected transconjugants were positive for the *mef(A)* gene as well as the 5' and 3' junctions of Tn1207.3-*comEC*. All amplicons were of the expected size. Thus, growth on erythromycin-selective plates can be taken as being strongly indicative of *mef(A)* acquisition by the previously susceptible, *mefA*-negative recipient strains and the integration of Tn1207.3 into the *comEC* locus.

The association of *emm* type with conjugative frequency is difficult to assess given the number of isolates of each *emm* type was low. However, the data show a trend for higher frequencies of conjugation among *emm1* and *emm4*, seen in the region of 10^{-6} to 10^{-7} with lower

Table 1 Strain characteristics and gene transfer frequencies of the *mef(A)*-bearing Tn1207.3 element (SMH036) to macrolide-susceptible *Strep. pyogenes*

<i>emm</i> type	Recipient	Multilocus sequence type	Site of isolation	Mean frequencies of three transfers per recipient \pm SEM
<i>emm1</i>	R366	ST28	Blood	$9.83 \times 10^{-6} \pm 1.87 \times 10^{-6}$
	R203	ST28	Blood	$1.13 \times 10^{-6} \pm 6.46 \times 10^{-7}$
	R057	ST28	Eye	$3.86 \times 10^{-6} \pm 1.91 \times 10^{-6}$
	R502	Not available	Blood	$4.23 \times 10^{-6} \pm 1.85 \times 10^{-6}$
<i>emm2</i>	R837	ST55	Blood	$3.17 \times 10^{-6} \pm 3.52 \times 10^{-7}$
	R007	ST55	Blood	$2.37 \times 10^{-6} \pm 5.30 \times 10^{-7}$
<i>emm3</i>	R032	ST315	Throat	$1.20 \times 10^{-7} \pm 1.52 \times 10^{-8}$
	R079	ST315	Throat	$9.80 \times 10^{-7} \pm 1.72 \times 10^{-7}$
<i>emm4</i>	R317	ST39	Blood	$4.50 \times 10^{-7} \pm 4.93 \times 10^{-8}$
	R211	ST39	Blood	$6.47 \times 10^{-7} \pm 1.05 \times 10^{-7}$
	R101	ST39	Ear	$6.17 \times 10^{-6} \pm 3.75 \times 10^{-7}$
<i>emm9</i>	R480	ST75	Blood	$9.20 \times 10^{-7} \pm 1.89 \times 10^{-7}$
	R097	ST75	Pus	$1.60 \times 10^{-7} \pm 1.53 \times 10^{-8}$
<i>emm11</i>	R098	ST22	Blood	$8.13 \times 10^{-7} \pm 1.67 \times 10^{-7}$
	R693	ST22	Blood	$3.67 \times 10^{-7} \pm 1.86 \times 10^{-8}$
<i>emm12</i>	R214	ST36	Blood	$1.19 \times 10^{-7} \pm 5.01 \times 10^{-8}$
	R113	ST36	Throat	$3.97 \times 10^{-7} \pm 4.97 \times 10^{-8}$
	R044	ST36	Throat	$6.00 \times 10^{-7} \pm 8.88 \times 10^{-8}$
<i>emm59</i>	R205	Not available	Blood	$4.36 \times 10^{-8} \pm 8.60 \times 10^{-9}$
<i>emm77</i>	R054	ST77	Skin	$<3.77 \times 10^{-8}$ (none detected)
	R115	ST77	Vagina	$<1.08 \times 10^{-8}$ (none detected)
<i>emm81</i>	R202	ST117	Blood	$3.97 \times 10^{-7} \pm 1.48 \times 10^{-7}$
<i>emm87</i>	R208	ST62	Blood	$7.20 \times 10^{-8} \pm 8.99 \times 10^{-9}$

rates seen particularly with the single isolates of *emm59* and *emm87*, in the region of 10^{-9} , Table 1. These patterns correlate well with the observed frequency of *mef(A)* among clinical isolates belonging to these *emm* types, *mef(A)* being highly prevalent among *emm1* and *emm4* isolates but having not been reported among *emm59* and *emm87* isolates (Creti *et al.* 2005; Grivea *et al.* 2006; Silva-Costa *et al.* 2008; Wajima *et al.* 2013). These data also highlight the potential for the emergence of *mef(A)*-mediated resistance in previously susceptible *emm* types. Two isolates, both belonging to *emm77*, did not produce transconjugants above the lower limit of detection, Table 1, suggesting that a low capacity for acquisition exists among *emm77*. This again correlates well with epidemiological data, and *emm77* has not previously been associated with *mef(A)*. Interestingly, previous work using transduction also failed to transfer *mefA* into an *emm 77* isolate (Di Luca *et al.* 2010). All seven *emm* types included here and reported previously to harbour the *mef(A)* gene were able to acquire it by conjugation *in vitro*. Taking only the *emm* types for which at least three isolates were included, *emm1*, *emm4* and *emm12*, the higher mean frequencies of transconjugants seen with *emm1* and *emm4* isolates were statistically significant compared to those seen with *emm12* isolates. This suggests that an association exists between *emm* type and the transfer frequency of Tn1207.3 but the testing of a larger collection of isolates is needed to explore this further.

Together the results demonstrate that significant variability exists in *mef(A)* gene conjugative transfer rates between *Strep. pyogenes* strains and that this phenomenon may contribute, at least in part, to the different frequencies of *mef(A)* observed among clinical GAS strains. The mechanisms responsible for these differing rates of *mefA* transfer are not known, but M proteins have previously been proposed to act as barriers for horizontal gene exchange and thus may play a role (Schmitz *et al.* 2003). Furthermore, we demonstrate potential for the development of *mef(A)*-mediated erythromycin resistance in currently susceptible lineages.

Materials and methods

Bacterial isolates

Group A streptococci isolates were collected, as part of a separate study, from two London Hospitals, St. Mary's and Paddington and Hammersmith, between 1993 and 2005 (McGregor and Spratt 2005). *mef(A)* recipient strains belonged to the following eleven *emm* types: *emm1*, *emm2*, *emm3*, *emm4*, *emm9*, *emm11*, *emm12*, *emm59*, *emm77*, *emm81* and *emm89*. Strains were chosen based on their availability for study and

because they include some of the most common lineages among clinical disease isolates in Europe (Steer *et al.* 2009) (Table 1). They also include lineages with observed differences in their frequency of *mef(A)* prevalence (Creti *et al.* 2005; Grivea *et al.* 2006; Silva-Costa *et al.* 2008; Wajima *et al.* 2013). Isolate SMH036 (*emm75*) was used as the Tn1207.3 donor and carries the Tn1207.3 element inserted into the *comEC* locus. Discrimination of *mef* gene subclasses A, E, O and I by RFLP analysis confirmed the presence of only *mef(A)* in SMH036. SMH036 was confirmed to be resistant to erythromycin with a MIC of 8 mg l⁻¹ and grew readily on 2 mg l⁻¹ erythromycin-selective blood plates. It was, however, susceptible to fusidic acid and rifampicin. All recipient strains were confirmed to be susceptible to erythromycin (MIC < 0.015–0.12 g l⁻¹) and unable to grow on selective blood agar plates containing 2 mg l⁻¹ erythromycin used in screening for transconjugants. The wild-type recipients were also confirmed to be susceptible to rifampicin and fusidic acid, and resistant derivatives of these, used to differentiate transconjugants from SMH036, were generated by inducing to spontaneous mutations conferring resistance to rifampicin or fusidic acid.

In vitro conjugative gene transfer of Tn1207.3

Filter mating was performed according to Giovanetti *et al.* (Giovanetti *et al.* 2002). Transconjugants were selected on blood agar plates containing either fusidic acid (25 mg l⁻¹) or rifampicin (25 mg l⁻¹) in addition to erythromycin (2 mg l⁻¹). Transfers were performed in triplicate.

Confirmation of *mef(A)* transfer

Putative transconjugants were tested for the presence of *mef(A)* by PCR using primers *mefF/mefR* as previously described (Sutcliffe *et al.* 1996b). Both Tn1207.3/*comEC* junctions were also examined by PCR, primers TnRjF/TnRjR (D'Ercole *et al.* 2005) spanning the 5' junction and *comECF/orfLF* (Santagati *et al.* 2003) spanning the 3' junction. Donor SMH036 was used as a positive control for these PCRs. Transconjugants were randomly selected (3–6 per donor/recipient combination) for PCR validation with a total of $n = 85$. All recipient strains produced no amplicons in these PCRs.

Acknowledgements

The support of Dr. Karen F. McGregor (University of West London) and the internal funding provided by the University of West London are gratefully acknowledged. EMH and GKP are supported by the MRC.

Conflict of interest

No conflict of interest declared.

References

- Banks, D.J., Porcella, S.F., Barbian, K.D., Martin, J.M. and Musser, J.M. (2003) Structure and distribution of an unusual chimeric genetic element encoding macrolide resistance in phylogenetically diverse clones of group A *Streptococcus*. *J Infect Dis* **188**, 1898–1908.
- Bley, C., Van Ded Linden, M. and Reinert, R.R. (2011) *mef(A)* is the predominant macrolide resistance determinant in *Streptococcus pneumoniae* and *Streptococcus pyogenes* in Germany. *Int J Antimicrob Agents* **37**, 425–431.
- Brenciani, A., Bacciaglia, A., Vignaroli, C., Pugnali, A., Varaldo, P.E. and Giovanetti, E. (2010) Phi m46.1, the main *Streptococcus pyogenes* element carrying *mef(A)* and *tet(O)* genes. *Antimicrob Agents Chemother* **54**, 221–229.
- Creti, R., Gherardi, G., Imperi, M., Von Hunolstein, C., Baldassarri, L., Pataracchia, M., Alfarone, G., Cardona, F. *et al.* (2005) Association of group A streptococcal *emm* types with virulence traits and macrolide-resistance genes is independent of the source of isolation. *J Med Microbiol* **54**, 913–917.
- D'Ercole, S., Peterelli, D., Prenna, M., Zampaloni, C., Catania, M.R., Ripa, S. and Vitali, L.A. (2005) Distribution of *mef(A)*-containing genetic elements in erythromycin-resistant isolates of *Streptococcus pyogenes* from Italy. *Clin Microbiol Infect* **11**, 927–930.
- Di Luca, M.C., D'Ercole, S., Peterelli, D., Prenna, M., Ripa, S. and Vitali, L.A. (2010) Lysogenic transfer of *mef(A)* and *tet(O)* genes carried by phi m46.1 among group A streptococci. *Antimicrob Agents Chemother* **54**, 4464–4466.
- Giovanetti, E., Brenciani, A., Burioni, R. and Varaldo, P.E. (2002) A novel efflux system in inducibly erythromycin-resistant strains of *Streptococcus pyogenes*. *Antimicrob Agents Chemother* **46**, 3750–3755.
- Giovanetti, E., Brenciani, A., Lupidi, R., Roberts, M.C. and Varaldo, P.E. (2003) Presence of the *tet(O)* gene in erythromycin- and tetracycline-resistant strains of *Streptococcus pyogenes* and linkage with either the *mef(A)* or the *erm(A)* gene. *Antimicrob Agents Chemother* **47**, 2844–2849.
- Grivea, I.N., AL-Lahham, A., Katopodis, G.D., Syrogiannopoulos, G.A. and Reinert, R.R. (2006) Resistance to erythromycin and telithromycin in *Streptococcus pyogenes* isolates obtained between 1999 and 2002 from Greek children with tonsillopharyngitis: phenotypic and genotypic analysis. *Antimicrob Agents Chemother* **50**, 256–261.
- McGregor, K.F. and Spratt, B.G. (2005) Identity and prevalence of multilocus sequence typing-defined clones of group A streptococci within a hospital setting. *J Clin Microbiol* **43**, 1963–1967.
- Santagati, M., Iannelli, F., Cascone, C., Campanile, F., Oggioni, M.R., Stefani, S. and Pozziz, G. (2003) The novel conjugative transposon Tn1207.3 carries the macrolide efflux gene *mef(A)* in *Streptococcus pyogenes*. *Microb Drug Resist* **9**, 243–247.
- Schmitz, F.J., Beyer, A., Charpentier, E., Normark, B.H., Schade, M., Fluit, A.C., Hafner, D. and Novak, R. (2003) Toxin-gene profile heterogeneity among endemic invasive European group A streptococcal isolates. *J Infect Dis* **188**, 1578–1586.
- Silva-Costa, C., Pinto, F.R., Ramirez, M., Melo-Cristino, J. and Portuguese Surveillance Group (2008) Decrease in macrolide resistance and clonal instability among *Streptococcus pyogenes* in Portugal. *Clin Microbiol Infect* **14**, 1152–1159.
- Steer, A.C., Law, I., Matatolu, L., Beall, B.W. and Carapetis, J.R. (2009) Global *emm* type distribution of group A streptococci: systematic review and implications for vaccine development. *Lancet Infect Dis* **9**, 611–616.
- Sutcliffe, J., Tait-Kamradt, A. and Wondrack, L. (1996a) *Streptococcus pneumoniae* and *Streptococcus pyogenes* resistant to macrolides but sensitive to clindamycin: a common resistance pattern mediated by an efflux system. *Antimicrob Agents Chemother* **40**, 1817–1824.
- Sutcliffe, J., Grebe, T., Tait-Kamradt, A. and Wondrack, L. (1996b) Detection of erythromycin-resistant determinants by PCR. *Antimicrob Agents Chemother* **40**, 2562–2566.
- Varaldo, P.E., Montanari, M.P. and Giovanetti, E. (2009) Genetic elements responsible for erythromycin resistance in streptococci. *Antimicrob Agents Chemother* **53**, 343–353.
- Wajima, T., Morozumi, M., Chiba, N., Shouji, M., Iwata, S., Sakata, H. and Ubukata, K. (2013) Associations of macrolide and fluoroquinolone resistance with molecular typing in *Streptococcus pyogenes* from invasive infections, 2010–2012. *Int J Antimicrob Agents* **42**, 447–449.